Association of Radiotherapy-Related Intestinal Injury and Cancer-related Fatigue: A Brief Review and Commentary

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> Radiotherapy treatment-induced intestinal injury and gut microbial perturbation/ dysbiosis have been implicated in the pathobiology of cancer-related fatigue. The objective of this brief review was to explore the available evidence of the relationship between intestinal injury and self-reported fatigue, especially among cancer patients. The scientific evidence—including our own—linking gut mucosal barrier dysfunction and gut microbial perturbation/dysbiosis induced by cancer treatment with worsening of cancer-related fatigue (perhaps through the gut-brain axis) is limited but promising. Emerging data suggest that lifestyle interventions and the administration of specific probiotics may favorably modulate the gut microbiota and potentially mediate beneficial effects leading to improvements in fatigue. [*P R Health Sci J 2021;40:6-11*]

Key words: Radiotherapy-related fatigue, Intestinal injury, Dysbiosis

Radiotherapy-related fatigue (RRF) is an unpleasant and distressing symptom experienced by approximately 67 to 74% of cancer patients (1,2), yet health care providers may under-recognize and undertreat it (3). RRF may also result in unwanted morbidities that have a great impact on healthrelated quality of life and the physical functioning of cancer survivors (4). A major gap in preventing and managing RRF is that its biological mechanisms have not been identified. One pivotal mechanism that may contribute to RRF development or exacerbation is the gut-brain axis.

Based on the renewed interest in the gut-brain axis and notable advances in high-dimensional sequencing techniques and in data analysis, the possible role of intestinal injury in the pathobiology of RRF is being further explored (5,6). More specifically, it has been proposed that mucosal barrier dysfunction and gut microbial dysbiosis induced by cancer treatment may contribute to the worsening of comorbidities, such as fatigue during a patient's pelvic radiotherapy (RT) (5,6,7,8). This brief review explores the available evidence of the relationship between intestinal injury and fatigue, especially among cancer patients.

Leaky-Gut

RT, producing a leaky gut (10,11). A leaky gut may facilitate the entry of large amounts of intestine-derived toxic factors (e.g., lipopolysaccharides, also called LPSs) into systemic circulation and may induce the aberrant activation of the immune system, resulting, for example, in cytokine-induced inflammatory reactions, which can affect brain function and increase symptom severity (5,7,8).

Serum levels of proteins released by dying mature enterocytes, such as fatty acid-binding protein (I-FABP) (12), as well as serum levels of citrulline (indicator of small intestine epithelial cell damage) (9) are among the proposed biomarkers of intestinal permeability (13). Additional information on the potential biomarkers of intestinal mucosal barrier injury can be found in recent extensive reviews (9,13). In fact, significant decreases in serum levels of citrulline and concentrations of I-FABP have been reported after myeloablative therapy

The authors have no conflict of interest to disclose.

Although the mechanisms underlying the structure and the regulation of the epithelial mucosal barrier are complex (9), evidence suggests that acute damage induced by RT treatments can result in gut mucosal barrier dysfunction or leaky gut, which in turn may be related to RRF. For example, evidence suggest that pelvic RT destroys intestinal integrity and increases intestinal permeability, which is greater at the end of

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(including chemotherapy and total body irradiation) in hematopoietic stem-cell transplant recipients (12). Plasma citrulline has also been linked to RT-induced small bowel mucosal atrophy and RRF (5,14). Wang et al. (2015) reported a significant negative correlation between citrulline concentration and fatigue scores at weeks 3 and 5 of pelvic RT. Jakobsson et al. (2010) reported similar findings. Although these findings suggest that the intestinal epithelial atrophy related to pelvic RT may contribute to RRF development or intensification, further exploration/understanding of the potential mechanisms by which leaky gut contributes to RRF is needed.

Endotoxemia

As previously highlighted, continuous damage to the gut mucosa by endogenous or exogenous events, such as RT treatment, often permit the translocation of bacteria and bacterial products (11,15). It has been suggested that in the event of the loss of the intestinal barrier, the commensal bacteria, such as indigenous microbiota, are no longer contained by the intestinal barrier (16). However, since the gut lumen is then predominantly exposed to gram-negative bacteria (containing elevated levels of endotoxins called LPSs), the commensal bacteria may enter the mesenteric lymph node and blood stream as a result of the barrier breach, causing endotoxemia (17). The increased translocation of LPSs from gram-negative bacteria may lead to autoantibody production, disruption of tight junctions, and both local gastrointestinal inflammation and systemic inflammation (17). Further, there is also evidence that the systemic increase of LPSs also contributes to central neuro-inflammation manifested by increased brain tumor necrosis factor alpha (TNF α) and by marked elevation of pro-inflammatory mediators (18). As a consequence, plasma levels of LPS and LPS-binding protein (LBP), as well as the LPS/LBP receptors CD14, are suggested endotoxemia markers (15).

Increased serum levels of LPS in pelvic cancer patients undergoing RT have been reported (14); however, there is evidence from an animal study linking LPS to fatigue (19). In one study, mice given LPSs (3 mg/kg) combined with swim stress (forced exercise to induce physical fatigue) showed decreased locomotor activity, decreased fall-off time in a rotarod test, and increased serum levels of interleukin-1 β and interleukin-6 (IL-6), compared with untreated mice (19). A number of investigators have attempted to show a relationship between endotoxemia markers and increased fatigue severity in other fatiguing conditions (e.g., chronic fatigue syndrome). For example, one study reported a significant positive correlation between IgA responses to LPSs of enterobacteria and selfreported fatigue in chronic fatigue syndrome patients (17). Future mechanistic studies exploring a causal link between endotoxemia and RRF, as well as mediation factors, may strengthen our proposition regarding the interplay of the gutbrain axis in the pathobiology of RRF.

Intestinal Dysbiosis

The gut is one of the organs of the body with the largest concentrations of bacteria, which range from 101 to 103 per gram in the upper intestine and from 1011 to 1012 per gram in the colon (9,20,21). Six major gut microbial phyla, namely Firmicutes (e.g., Clostridium genus), Bacteroidetes (e.g., Prevotella and Bacteroides genera), Actinobacteria (e.g., Bifidobacterium genus), Proteobacteria (e.g., Escherichia and Shigella genera), Fusobacteria, and Verrucomicrobia, compose the gut bacterial microbiome of healthy individuals (21). Furthermore, perturbation of the composition of the gut bacteria (e.g., caused by various factors, such as RT and/ or chemotherapy, stress, exercise, dietary pattern, antibiotic treatment) may result in gut microbial dysbiosis (6,9,11,22). Gut microbial dysbiosis is often characterized by lower species diversity, fewer beneficial microbes, the expansion of pathogenic microbes, and the production of factors toxic or harmful to host cells (22).

Culture-based and 16S ribosomal RNA (rRNA) gene sequencing studies of stool bacteria have revealed some evidence of a general decrease in the biomass of intestinal microbiota and an imbalance of the intestine bacterial community towards decreased numbers of commensal bacteria associated to RT (11,14,23). For example, a small study (n = 10) of abdominal cancer patients, using DNA fingerprinting and cloningsequencing techniques, found that patients with diarrhea had significantly modified bacterial profiles during pelvic RT compared both to healthy controls and to patients with no diarrhea but who received RT (23). Additionally, patients with diarrhea exhibited increased levels of Actinobacteria, increased levels of Bacilli, and decreased levels of Clostridia compared to those patients with no diarrhea. Indeed, the overgrowth of gram-negative Bacilli has been linked to the pathogenesis of intestinal radiation toxicity (23). Similarly, another small study, this one of gynecological cancer patients undergoing RT and using pyrosequencing analysis of the 16S rRNA gene, showed a significant decrease in phylum Firmicutes and an increase in phylum Fusobacterium in the participants compared to healthy volunteers (24). At the family level, Clostridiaceae and Eubacteriaceae were more common and Prevotellaceae, Oscillospiraceae, and Fusobacteriaceae less common in cancer patients compared to healthy individuals. Importantly, a reduction in the diversity of the Firmicutes phylum has been linked to some systemic inflammatory diseases (e.g., bowel disorders) (22,23,25). However, the commensal intestinal microbiota might also have beneficial effects. For example, in one animal study, the administration of Prevotella histicola, native to the human gut, was linked to immunomodulatory and anti-inflammatory capabilities (26). In addition, a significant decrease in the Shannon diversity index (bacterial richness and evenness) during pelvic RT has been reported (14). Nonetheless, these findings need to be interpreted with caution, as the previously mentioned studies included several important limitations (e.g., small sample size, limited number of sequences analyzed, exclusion of additional changes in other groups of bacteria). It is also possible that these changes in the microbiota profile may be attributed to the diagnosed cancer (11). For example, *Fusobacterium nucleatum* infection has been found to be associated with colorectal cancer development, with an increased risk for metastasis, and with a potential colorectal cancer biomarker (27).

Interestingly, although there was a paucity of studies linking imbalance of the intestinal microbiota, known as dysbiosis, with RRF, one of the reviewed studies suggested a potential relationship between dysbiosis and fatigue interference during pelvic radiation therapy (14). Wang et al.'s 2015 study did not report any direct associations between gut microbial dysbiosis and fatigue scores using the Multidimensional Fatigue Inventory. However, the latter study found that microbial diversity and richness and the Firmicutes/Bacteroides ratio were significantly altered prior to pelvic radiation therapy in patients who later developed diarrhea and reported significant worsening in fatigue. Although bacterial communities have been the primary focus of gut dysbiosis research, recently it has been suggested that fungal dysbiosis (28) may have a role in the experience of fatigue (29). One small study using 18S rRNA sequencing found that healthy controls (n = 17) had a slightly lower average relative abundance of fungi compared to patients with myalgic encephalomyelitis or chronic fatigue syndrome (n = 17) (30.6% vs. 33.6%) (29). The latter study also reported reduced gut eukaryotic diversity (eukaryotes in the gut include protozoa and fungi), although non-statistically significant, among myalgic encephalomyelitis/chronic fatigue syndrome patients. In addition, the authors of the latter study highlighted limitations in eukaryotic research, including the low abundance and diversity of eukaryotes in the human gut microbiome, the limited number of high-quality eukaryotic reads, and the possibility that 18S databases were incomplete at that moment (29). Despite this evidence, not enough comprehensive 16S rRNA and/or 18S rRNA gene sequencing data exists to determine whether RRF is associated with RTinduced changes of the gut microbiota. Such findings could broaden our understanding of the pathobiology linking intestinal dysbiosis and RRF.

Metabolites

Recent interest in the possibility that physiological levels of numerous bioactive metabolic products (metabolites) formed by the gut microbiota may be associated with cancer-related symptoms has emerged. Indeed, pre-clinical studies have shown significant increases of metabolites, such as valine, leucine, isoleucine, and 2-oxoisopentanoate, in plasma, as well as significant decreases in citrulline and hydroxyproline in a rat model of fatigue compared to the control and foodrestricted groups (30). Additionally, one pilot clinical study of breast cancer patients undergoing chemotherapy showed that increased fatigue, post-chemotherapy, was associated with decreased and/or increased levels of metabolites from the tryptophan pathway (31). Increased fatigue was associated with decreased blood levels of multiple bile-related compounds (e.g., taurochenodeoxycholic acid, taurocholic acid) and with increased levels of other classes of metabolites, such as diazines, monosaccharides, and fatty acids (31). The latter study also found significant associations between other metabolites and other symptoms, including pain and depression. There is also evidence of increased d-lactic acid (a product of the microbial fermentation of carbohydrate) and urine metabolites (e.g., amino-hydroxy-N-methyl-pyrrolidine) in patients with other fatiguing and cognitive-impairing conditions, such as chronic fatigue syndrome (32,33). Moving forward, metabolomic studies may enable the identification of metabolites or metabolic pathways that may contribute to the understanding of the pathobiology of fatigue and identify potential targets to develop interventions for RRF (30,31).

Inflammation

Cancer and its treatments affect many biological networks, including those related to the immune and inflammation responses. Immune activation is frequently characterized by an elevation of pro-inflammatory cytokines, an increased expression of T lymphocyte, and the decreased function of natural killer (NK) cells. For example, RT was found to induce the elevation of circulating levels of pro-inflammatory cytokines (TNFa, IL-6, CD8+ T lymphocytes, and NK cells in patients with prostate cancer [34]). Pro-inflammatory cytokines, such as interleukin-1, interleukin-1 receptor antagonist, and IL-6, have been associated with fatigue severity in cancer survivors (35). High levels of c-reactive protein and haptoglobin have been shown to significantly correlate with fatigue (5). Immune dysregulation can also be triggered by the translocation of microbes from an inflamed and/or leaky-gut (15) as a side effect of cancer therapy.

There is also evidence that the gut microbiome regulates important processes, including inflammation and immunity, at both the local gastrointestinal and the systemic inflammation levels (22). Evidence from a number of animal studies suggests that signals from commensal bacteria deliver tonic immune stimulation, which in turn results in the activation threshold of innate antiviral immunity (36). Additionally, commensal microorganisms may regulate gut inflammatory responses. Some Bifidobacterium strains and Bacteroides thetaiotaomicron strains are able to attenuate inflammation via the suppression of nuclear factor kappa B, resulting in the decrease of proinflammatory cytokines and cyclooxygenase (24,37). Similarly, microorganisms in Clostridium cluster XIVa have been reported to play a role in decreasing the inflammation of the gut epithelium (24,38). Furthermore, there is also evidence that chemotherapy induced increases in Clostridium cluster XI and Enterobacteriaceae abundance to potentially pathogenic levels may be associated with diarrhea (e.g., Clostridium difficileassociated diarrhea) (39). Notably, further comprehensive molecular analysis studies are needed to confirm and determine

both the role of a certain group of bacteria in the etiology of RT-induced dysbiosis and the side effects of RT (e.g., mucositis and diarrhea).

Implications for practice

RRF continues to be a prevalent, debilitating, and complex symptom experienced by many cancer patients. Since the findings of the current brief review suggest that cancer treatments have significant consequences on intestinal permeability and gut dysbiosis, which consequences may include fatigue, it is plausible that pharmacological (e.g., probiotics) and lifestyle interventions targeting the modulation of the gut microbiota may provide insight into the prevention or amelioration of fatigue (40,41). For example, one study found that 12 weeks of probiotic supplementation (a commercial combination of Lactobacillus rhamnosus and Lactobacillus helveticus) improved the fatigue, depression, mood, and quality of life of colorectal cancer survivors, with such improvements not being observed in the members of the placebo group (42). Another study examined associations between changes in gut microbiota composition and changes in cardiorespiratory fitness, fatigue, anxiety, depression, and sleep dysfunction in breast cancer survivors before and 3 months after participating in a physical activity trial (43). This latter study found that increased fatigue interference mean scores are associated with an increased mean Shannon diversity index (43). In addition, that study showed that the magnitude of change in fatigue interference was associated with the frequency of Faecalibacterium and Prevotella genera. This is important because there is increasing evidence of the potential anti-inflammatory roles of Prevotella histicola and Faecalibacterium prausnitzii (26,44).

It is also possible that strategies aimed at restoring and maintaining a healthy intestinal barrier (e.g., avoiding a high fat diet and processed food, eating meals rich in fiber and fruits) (9) could be introduced into a nutritional intervention study that might, upon completion, provide insight into the prevention or amelioration of fatigue. Clinical trials designed to explore the efficacy of diets rich in fruits, vegetables, whole grains, and omega-3 fatty acid-rich foods in reducing fatigue have reported improvements in fatigue and sleep among breast cancer survivors (45). Additionally, better-quality diets (as measured by the Healthy Eating Index 2010) have been found to be associated with lower fatigue during cancer survivorship (46). In contrast, one study of prostate cancer patients undergoing RT found that reducing the intake of insoluble dietary fibers and lactose had no effect on either fatigue or other aspects of quality of life (47). Much research exploring the clinical relevance of promising pharmacological and lifestyle interventions is needed.

Research implications, including our preliminary gut microbiome work

Pilot studies are underway, including our own, to explore whether RT-induced dysbiosis is associated with the symptom experience of rectal cancer patients undergoing pelvic RT. Using Illumina MiSeq technology, we recently conducted a 16S ribosomal RNA (rRNA) gene sequencing pilot study from stool samples of rectal cancer patients undergoing chemoradiation. This was done to provide initial evidence of the biological/gut microbial processes related to the associations of chemoradiation and dysbiosis with the symptom experience in this population. In fact, we recently reported the association between self-reported fatigue (48) or sleepdisturbance (49) and gut microbial alpha diversity during CRT, as well as significant correlations between the abundances of Bacteroides and Blautia2 and co-occurring symptoms at the end of chemoradiation (50). However, because of the limited number of participants and lack of a control/comparison group, our results were exploratory in nature and should be interpreted with caution. Small sample sizes present additional challenges for statistical analyses in terms of significance testing after multiple corrections and the inability to control for multiple important covariates (e.g., age, sex assigned at birth, body mass index). Future studies using metagenomic and metatranscriptomic approaches, and/or using real-time PCR to quantify certain bacteria beyond the genus level, may further validate our findings. Studies examining the strength of the associations among patient (host) characteristics, gut dysbiosis, intestinal injury, and host inflammatory markers with fatigue might be part of our future research. Gaining this knowledge may strengthen our proposition regarding the interplay of the gut-brain axis in the pathobiology of RRF.

Moving forward, despite notable milestones already having been reached in the field of microbiome research, the following important challenges remain: the development of novel bioinformatic and novel molecular tools, the development of methods for integrating new and current data, and the generation of data that are directly proportional to their biological significance, among others (51). Addressing these challenges will allow for the transformation of a chaotic plethora of metagenomic information and physiologically relevant material into manageable bites of biological data, thereby advancing the understanding of both symptom science and personalized cancer care.

Conclusion

The pathobiology of RRF is not completely understood but is considered multifactorial, with the gut–brain axis, mucosal barrier dysfunction, and gut microbial dysbiosis thought to play roles. Identifying patients at a greater risk of developing RRF may promote the optimized delivery of anticancer therapy and the provision of tailored supportive care and aid all involved in making well-informed, evidence-based decisions regarding treatment. Recent evidence suggests that gut microbiota– targeted interventions (e.g., probiotics) may potentially modulate pelvic radiation–induced side effects (52). However, because microbiota differentiation can be observed in each individual, personalized plans of care and therapeutic strategies should be the focus of future personalized cancer care research (53,54). Larger-scale longitudinal studies are needed to further investigate the roles of other factors (e.g., health status, diet, body mass index, lifestyle, social factors, environmental factors, pain, stress, diarrhea) that may affect the variability of gut microbiota measurements and/or fatigue symptoms in patients with cancer who are receiving RT.

Resumen

La lesión intestinal inducida por el tratamiento de radioterapia y la perturbación microbiana intestinal/disbiosis se han implicado en la patobiología de la fatiga relacionada al cáncer. El objetivo de esta breve revisión fue explorar la evidencia disponible de la relación entre la lesión intestinal y el auto reporte de la fatiga, especialmente entre los pacientes con cáncer. La evidencia científica, incluyendo la nuestra, que vincula la disfunción de la barrera mucosa intestinal y la perturbación/disbiosis microbiana intestinal inducida por el tratamiento del cáncer con el empeoramiento de la fatiga, tal vez a través del eje intestino-cerebro, es limitada, pero prometedora. Los datos emergentes sugieren que intervenciones en el estilo de vida y la administración de probióticos específicos podrían modular de manera favorable la microbiota intestinal y potencialmente mediar efectos beneficiosos, resultando en mejoras en la fatiga.

Acknowledgments

This article was made possible by the National Institute of Nursing Research (NINR) of the National Institutes of Health (NIH) under award number F32NR016618 and University of Puerto Rico (UPR) NIH grant awards 2U54MD007587 and CA096297/CA096300. The content is solely the responsibility of the authors and does not necessarily represent the official views of the NIH.

References

- Langston B, Armes J, Levy A, Tidey E, Ream E. The prevalence and severity of fatigue in men with prostate cancer: a systematic review of the literature. Support Care Cancer. 2013;21(6):1761-1771. doi:10.1007/ s00520-013-1751-5
- O'Gorman C, Denieffe S, Gooney M. Literature review: preoperative radiotherapy and rectal cancer - impact on acute symptom presentation and quality of life. J Clin Nurs. 2014;23(3-4):333-351. doi:10.1111/ jocn.12138
- Yoon J, Malin JL, Tisnado DM, et al. Symptom management after breast cancer treatment: is it influenced by patient characteristics?. Breast Cancer Res Treat. 2008;108(1):69-77. doi:10.1007/s10549-007-9580-1
- Gonzalez VJ, McMillan S, Pedro E, Tirado-Gomez M, Saligan LN. The Health related Quality of Life of Puerto Ricans during Cancer Treatments; A Pilot Study. P R Health Sci J. 2018;37(1):46-51.
- Jakobsson S, Ahlberg K, Taft C, Ekman T. Exploring a link between fatigue and intestinal injury during pelvic radiotherapy. Oncologist. 2010;15(9):1009-1015. doi:10.1634/theoncologist.2010-0097.
- Jordan KR, Loman BR, Bailey MT, Pyter LM. Gut microbiota-immunebrain interactions in chemotherapy-associated behavioral comorbidities. Cancer. 2018;124(20):3990-3999. doi:10.1002/cncr.31584

- Ferreira MR, Muls A, Dearnaley DP, Andreyev HJ. Microbiota and radiation-induced bowel toxicity: lessons from inflammatory bowel disease for the radiation oncologist. Lancet Oncol. 2014;15(3):e139-e147. doi:10.1016/S1470-2045(13)70504-7
- Kiecolt-Glaser JK, Derry HM, Fagundes CP. Inflammation: depression fans the flames and feasts on the heat. Am J Psychiatry. 2015;172(11):1075-1091. doi:10.1176/appi.ajp.2015.15020152
- Kelly JR, Kennedy PJ, Cryan JF, Dinan TG, Clarke G, Hyland NP. Breaking down the barriers: the gut microbiome, intestinal permeability and stress-related psychiatric disorders. Front Cell Neurosci. 2015;9:392. Published 2015 Oct 14. doi:10.3389/fncel.2015.00392
- Nejdfors P, Ekelund M, Weström BR, Willén R, Jeppsson B. Intestinal permeability in humans is increased after radiation therapy. Dis Colon Rectum. 2000;43(11):1582-1588. doi:10.1007/BF02236743
- Touchefeu Y, Montassier E, Nieman K, et al. Systematic review: the role of the gut microbiota in chemotherapy- or radiation-induced gastrointestinal mucositis - current evidence and potential clinical applications. Aliment Pharmacol Ther. 2014;40(5):409-421. doi:10.1111/apt.12878
- Derikx JP, Blijlevens NM, Donnelly JP, et al. Loss of enterocyte mass is accompanied by diminished turnover of enterocytes after myeloablative therapy in haematopoietic stem-cell transplant recipients. Ann Oncol. 2009;20(2):337-342. doi:10.1093/annonc/mdn579
- Kelly JR, Kennedy PJ, Cryan JF, Dinan TG, Clarke G, & Hyland NP. Breaking down the barriers: the gut microbiome, intestinal permeability and stress-related psychiatric disorders. Frontiers in cellular neuroscience. 2015;9, 392. ttps://doi.org/10.3389/fncel.2015.00392
- Wang A, Ling Z, Yang Z, et al. Gut microbial dysbiosis may predict diarrhea and fatigue in patients undergoing pelvic cancer radiotherapy: a pilot study. PLoS One. 2015;10(5):e0126312. Published 2015 May 8. doi:10.1371/journal.pone.0126312
- Giloteaux L, Goodrich JK, Walters WA, Levine SM, Ley RE, Hanson MR. Reduced diversity and altered composition of the gut microbiome in individuals with myalgic encephalomyelitis/chronic fatigue syndrome. Microbiome. 2016;4(1):30. Published 2016 Jun 23. doi:10.1186/s40168-016-0171-4
- Tlaskalová-Hogenová H, Stepánková R, Hudcovic T, et al. Commensal bacteria (normal microflora), mucosal immunity and chronic inflammatory and autoimmune diseases. Immunol Lett. 2004;93(2-3):97-108. doi:10.1016/j.imlet.2004.02.005
- Maes M, Twisk FN, Kubera M, Ringel K, Leunis JC, Geffard M. Increased IgA responses to the LPS of commensal bacteria is associated with inflammation and activation of cell-mediated immunity in chronic fatigue syndrome. J Affect Disord. 2012;136(3):909-917. doi:10.1016/j. jad.2011.09.010
- Qin L, Wu X, Block ML, et al. Systemic LPS causes chronic neuroinflammation and progressive neurodegeneration. Glia. 2007;55(5):453-462. doi:10.1002/glia.20467
- Zhang ZT, Du XM, Ma XJ, et al. Activation of the NLRP3 inflammasome in lipopolysaccharide-induced mouse fatigue and its relevance to chronic fatigue syndrome. J Neuroinflammation. 2016;13(1):71. Published 2016 Apr 5. doi:10.1186/s12974-016-0539-1
- O'Hara AM, Shanahan F. The gut flora as a forgotten organ. EMBO Rep. 2006;7(7):688-693. doi:10.1038/sj.embor.7400731
- Rinninella E, Raoul P, Cintoni M, et al. What is the Healthy Gut Microbiota Composition? A Changing Ecosystem across Age, Environment, Diet, and Diseases. Microorganisms. 2019;7(1):14. Published 2019 Jan 10. doi:10.3390/microorganisms7010014
- Bajic JE, Johnston IN, Howarth GS, Hutchinson MR. From the Bottom-Up: Chemotherapy and Gut-Brain Axis Dysregulation. Front Behav Neurosci. 2018;12:104. Published 2018 May 22. doi:10.3389/fnbeh.2018.00104
- Manichanh C, Varela E, Martinez C, et al. The gut microbiota predispose to the pathophysiology of acute postradiotherapy diarrhea. Am J Gastroenterol. 2008;103(7):1754-1761. doi:10.1111/j.1572-0241.2008.01868.x
- 24. Nam YD, Kim HJ, Seo JG, Kang SW, Bae JW. Impact of pelvic radiotherapy on gut microbiota of gynecological cancer patients revealed by massive

pyrosequencing. PLoS One. 2013;8(12):e82659. Published 2013 Dec 18. doi:10.1371/journal.pone.0082659

- Frank DN, St Amand AL, Feldman RA, Boedeker EC, Harpaz N, Pace NR. Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases. Proc Natl Acad Sci U S A. 2007;104(34):13780-13785. doi:10.1073/pnas.0706625104
- Marietta EV, Murray JA, Luckey DH, et al. Suppression of Inflammatory Arthritis by Human Gut-Derived Prevotella histicola in Humanized Mice. Arthritis Rheumatol. 2016;68(12):2878-2888. doi:10.1002/art.39785
- Peng BJ, Cao CY, Li W, et al. Diagnostic Performance of Intestinal Fusobacterium nucleatum in Colorectal Cancer: A Meta-Analysis. Chin Med J (Engl). 2018;131(11):1349-1356. doi:10.4103/0366-6999.232814
- Iliev ID, Leonardi I. Fungal dysbiosis: immunity and interactions at mucosal barriers. Nat Rev Immunol. 2017;17(10):635-646. doi:10.1038/ nri.2017.55
- Mandarano AH, Giloteaux L, Keller BA, Levine SM, Hanson MR. Eukaryotes in the gut microbiota in myalgic encephalomyelitis/chronic fatigue syndrome. PeerJ. 2018;6:e4282. Published 2018 Jan 22. doi:10.7717/ peerj.4282
- Kume S, Yamato M, Tamura Y, et al. Potential biomarkers of fatigue identified by plasma metabolome analysis in rats. PLoS One. 2015;10(3):e0120106. Published 2015 Mar 20. doi:10.1371/journal. pone.0120106
- Lyon DE, Starkweather A, Yao Y, et al. Pilot Study of Metabolomics and Psychoneurological Symptoms in Women With Early Stage Breast Cancer. Biol Res Nurs. 2018;20(2):227-236. doi:10.1177/1099800417747411
- Galland L. The gut microbiome and the brain. J Med Food. 2014;17(12):1261-1272. doi:10.1089/jmf.2014.7000
- Sheedy JR, Wettenhall RE, Scanlon D, et al. Increased d-lactic Acid intestinal bacteria in patients with chronic fatigue syndrome. In Vivo. 2009;23(4):621-628.
- Hurwitz MD, Kaur P, Nagaraja GM, Bausero MA, Manola J, Asea A. Radiation therapy induces circulating serum Hsp72 in patients with prostate cancer. Radiother Oncol. 2010;95(3):350-358. doi:10.1016/j. radonc.2010.03.024
- 35. Rich T, Zhao F, Cruciani RA, Cella D, Manola J, Fisch MJ. Association of fatigue and depression with circulating levels of proinflammatory cytokines and epidermal growth factor receptor ligands: a correlative study of a placebo-controlled fatigue trial. Cancer Manag Res. 2017;9:1-10. Published 2017 Jan 31. doi:10.2147/CMAR.S115835
- Abt MC, Osborne LC, Monticelli LA, et al. Commensal bacteria calibrate the activation threshold of innate antiviral immunity. Immunity. 2012;37(1):158-170. doi:10.1016/j.immuni.2012.04.011
- Khokhlova EV, Smeianov VV, Efimov BA, Kafarskaia LI, Pavlova SI, Shkoporov AN. Anti-inflammatory properties of intestinal Bifidobacterium strains isolated from healthy infants. Microbiol Immunol. 2012;56(1):27-39. doi:10.1111/j.1348-0421.2011.00398.x
- Matsumoto M, Benno Y. The relationship between microbiota and polyamine concentration in the human intestine: a pilot study. Microbiol Immunol. 2007;51(1):25-35. doi:10.1111/j.1348-0421.2007.tb03887.x
- Lin XB, Dieleman LA, Ketabi A, et al. Irinotecan (CPT-11) chemotherapy alters intestinal microbiota in tumour bearing rats. PLoS One. 2012;7(7):e39764. doi:10.1371/journal.pone.0039764
- Minton O, Stone P, Richardson A, Sharpe M, Hotopf M. Drug therapy for the management of cancer related fatigue. Cochrane Database Syst Rev. 2008;(1):CD006704. Published 2008 Jan 23. doi:10.1002/14651858. CD006704.pub2

- Mustian KM, Alfano CM, Heckler C, et al. Comparison of Pharmaceutical, Psychological, and Exercise Treatments for Cancer-Related Fatigue: A Meta-analysis. JAMA Oncol. 2017;3(7):961-968. doi:10.1001/jamaoncol.2016.6914
- 42. Lee JY, Chu SH, Jeon JY, et al. Effects of 12 weeks of probiotic supplementation on quality of life in colorectal cancer survivors: a double-blind, randomized, placebo-controlled trial. Dig Liver Dis. 2014;46(12):1126-1132. doi:10.1016/j.dld.2014.09.004
- 43. Paulsen JA, Ptacek TS, Carter SJ, et al. Gut microbiota composition associated with alterations in cardiorespiratory fitness and psychosocial outcomes among breast cancer survivors. Support Care Cancer. 2017;25(5):1563-1570. doi:10.1007/s00520-016-3568-5
- 44. Sitkin S, Pokrotnieks J. Clinical Potential of Anti-inflammatory Effects of Faecalibacterium prausnitzii and Butyrate in Inflammatory Bowel Disease. Inflamm Bowel Dis. 2019;25(4):e40-e41. doi:10.1093/ibd/izy258
- Zick SM, Colacino J, Cornellier M, Khabir T, Surnow K, Djuric Z. Fatigue reduction diet in breast cancer survivors: a pilot randomized clinical trial. Breast Cancer Res Treat. 2017;161(2):299-310. doi:10.1007/s10549-016-4070-y
- 46. George SM, Alfano CM, Neuhouser ML, et al. Better postdiagnosis diet quality is associated with less cancer-related fatigue in breast cancer survivors. J Cancer Surviv. 2014;8(4):680-687. doi:10.1007/s11764-014-0381-3
- 47. Pettersson A, Johansson B, Persson C, Berglund A, Turesson I. Effects of a dietary intervention on acute gastrointestinal side effects and other aspects of health-related quality of life: a randomized controlled trial in prostate cancer patients undergoing radiotherapy. Radiother Oncol. 2012;103(3):333-340. doi:10.1016/j.radonc.2012.04.006
- González-Mercado VJ, Pérez-Santiago J, Lyon D, et al. The Role of Gut Microbiome Perturbation in Fatigue Induced by Repeated Stress from Chemoradiotherapy: A Proof of Concept Study. Adv Med. 2020;2020:6375876. Published 2020 Feb 7. doi:10.1155/2020/6375876
- González-Mercado VJ, Sarkar A, Penedo FJ, et al. Gut microbiota perturbation is associated with acute sleep disturbance among rectal cancer patients. J Sleep Res. 2020;29(3):e12915. doi:10.1111/jsr.12915
- González-Mercado VJ, Henderson WA, Sarkar A, et al. Changes in Gut Microbiome Associated With Co-Occurring Symptoms Development During Chemo-Radiation for Rectal Cancer: A Proof of Concept Study [published online ahead of print, 2020 Jul 23]. Biol Res Nurs. 2020;1099800420942830. doi:10.1177/1099800420942830
- Alves LF, Westmann CA, Lovate GL, de Siqueira GMV, Borelli TC, Guazzaroni ME. Metagenomic Approaches for Understanding New Concepts in Microbial Science. Int J Genomics. 2018;2018:2312987. Published 2018 Aug 23. doi:10.1155/2018/2312987
- Shibata N, Kunisawa J, Kiyono H. Dietary and Microbial Metabolites in the Regulation of Host Immunity. Front Microbiol. 2017;8:2171. Published 2017 Nov 7. doi:10.3389/fmicb.2017.02171
- 53. Deng X, Li Z, Li G, Li B, Jin X, Lyu G. Comparison of Microbiota in Patients Treated by Surgery or Chemotherapy by 16S rRNA Sequencing Reveals Potential Biomarkers for Colorectal Cancer Therapy. Front Microbiol. 2018;9:1607. Published 2018 Jul 17. doi:10.3389/fmicb.2018.01607
- 54. Wardill HR, Tissing WJE. Determining risk of severe gastrointestinal toxicity based on pretreatment gut microbial community in patients receiving cancer treatment: a new predictive strategy in the quest for personalized cancer medicine. Curr Opin Support Palliat Care. 2017;11(2):125-132. doi:10.1097/SPC.00000000000265