
A novel technique leading to complete sensory and motor recovery across a long peripheral nerve gap

ONIX REYES, MD*; IVAN J. SOSA, MD†; JOSÉ SANTIAGO, MD‡; DAMIEN P. KUFFLER, Ph D**

Sensory nerve grafts are the “gold standard” for inducing neurological recovery in peripheral nerves with a gap. However, the effectiveness of sensory nerve grafts is variable, generally not leading to complete sensory and motor recovery, with good recovery limited to gaps shorter than 2 cm, and the extent of recovery decreasing with increasing graft length. An alternative technique using a conduit filled with pure fibrin to bridge a nerve gap leads to only limited neurological recovery. We tested the effectiveness of a novel nerve repair technique in which a 5-cm long radial nerve gap

was repaired using two sural nerve graft surrounded by a collage tube filled with pure fibrin. By 1½ years post surgery, the patient recovered complete sensory and motor function. In conclusion, this study suggests that the combination of pure fibrin surrounding sural nerve grafts is responsible for inducing the extensive neurological recovery induced by either pure fibrin or sural grafts alone. This technique is presently being tested in a clinical trial.

Key words: Axon regeneration, Fibrin, Sural nerve graft.

The standard clinical technique for repairing a peripheral nerve gap when both ends can not be anastomosed is to bridge the gap with autologous nerve grafts harvested from the cutaneous saphenous or sural nerves (1-6). The assumption is that sensory nerve Schwann cell-released neurotropic factors and molecules in the nerve extracellular matrix should promote and direct axon regeneration across the nerve gap (7-12).

Sensory nerve grafts result in some neurological recovery in about 86% of patients with nerve gaps 1-5-cm in length, with generally limited recovery for gaps 1.5-2 cm in length, very limited recovery for gaps longer than 2 cm in length, and decreasing with further increases in gap length (4, 6, 10, 13-18). The reason for the limited recovery appears to be that the sensory axons use the sural nerve grafts as a passive scaffold across which the axons regenerate, rather than as an active regeneration-promoting pathway (4, 19, 20).

Vascularized sensory nerve grafts significantly improve the extent of neurological recovery compared to that induced by non-vascularized nerve grafts (21, 22). However, the far more extensive surgery required to prepare vascularized nerve grafts, significantly limits their general application.

Alternative methods for bridging nerve gaps are pseudo nerves, CNS tissue grafts, collagen guides, guides filled with neurotrophic factors, gradients of factors, antibodies, factors that induce inflammation, biodegradable polymer tubes, and filling tubes with various materials such as collagen or artificial fibrin matrix (5, 18, 23-35). However, most of these techniques fail to promote axon regeneration across gaps longer than 2 cm and provide no better neurological recovery than sural nerve grafts (35, 36). Therefore, sensory nerve grafts use remain the “gold standard” for the clinical repair of transected peripheral nerves (5, 37, 38).

Case Report

The present surgical repair was performed to determine whether a modification of the standard sural nerve graft technique might lead to more extensive neurological recovery than typically seen across a long radial nerve gap.

A 21-year-old male presented to the hospital two months after a traumatic radial nerve lesion at the elbow. Exposure of the injury site showed both nerve stumps to be inflamed (Figure 1A). After removing the damaged tissue, the nerve had a 5-cm long gap inflamed (Figure 1B). Two lengths of sural nerve were placed into the gap and sutured in place in accordance with standard surgical repair procedures inflamed (Figure 1C-D). A sheet of collagen (artificial dura, Duraguard, Synovis Corp. St. Paul, MN, USA) was sewn into a tube around the grafts, with a diameter slightly larger than that of the radial nerve inflamed. The ends of the

*Doctor's Center Hospital, Manati, PR, †Section of Neurological Surgery, Medical Sciences Campus, UPR, ‡Department of Orthopedic Surgery, Medical Sciences Campus, UPR, **Institute of Neurobiology, Medical Sciences Campus, UPR

Address correspondence to: Damien Kuffler, Ph D, Institute of Neurobiology, Medical Sciences Campus, University of Puerto Rico, 201 Blvd. del Valle, San Juan, PR 00901, Tel: (787) 721-1235 FAX: (787) 725-1289 e-mail: dkuffler@hotmail.com

radial nerve were inserted about 3 mm into the collagen tube, and the collagen tube was sewn to the epineurium of the ends of the radial nerve (Figure 1E). The space between the nerve grafts and tube was then filled with fibrin (cryoprecipitate) combined with thrombin. This combination led to the polymerization of fibrinogen within the cryoprecipitate into a 3-dimensional matrix (Figure 1F).

By 1½ years post surgery, the patient showed no signs of muscle atrophy, had recovered complete motor and sensory function and suffers no pain related to the nerve injury and repair, and had had no adverse effects from the repair. The patient developed sensitivity to touch, temperature, pain, vibration and normal 2-point discrimination on the dorsal aspect of his hand, identical to that of his un-injured hand.

Motor recovery included full and graded wrist and finger extension, indicating the establishment of many motor

units. The force exerted by the wrist and fingers was virtually identical to that of the uninjured hand, indicating full muscle innervation. Graded electrical stimulation of the radial nerve above the lesion site induced graded wrist and finger extension,

Evoked potentials were recorded when the radial nerve was stimulated from the thumb notch, forearm, elbow and above the elbow (at distances of 14-22 cm). The signal latency was normal at 0.2 ms, with an amplitude of 32 µv. Needle recordings from finger extensor muscles showed the muscles to be electrically silent during periods of muscle inactivity, and with no fibrillations. Voluntarily movement of selected fingers gave rise to normal evoked motor activity for the appropriate finger. These results indicate normal muscle innervation by the regenerated radial nerve axons and normal evoked action potentials.

The patient passed a rigorous military physical exam,

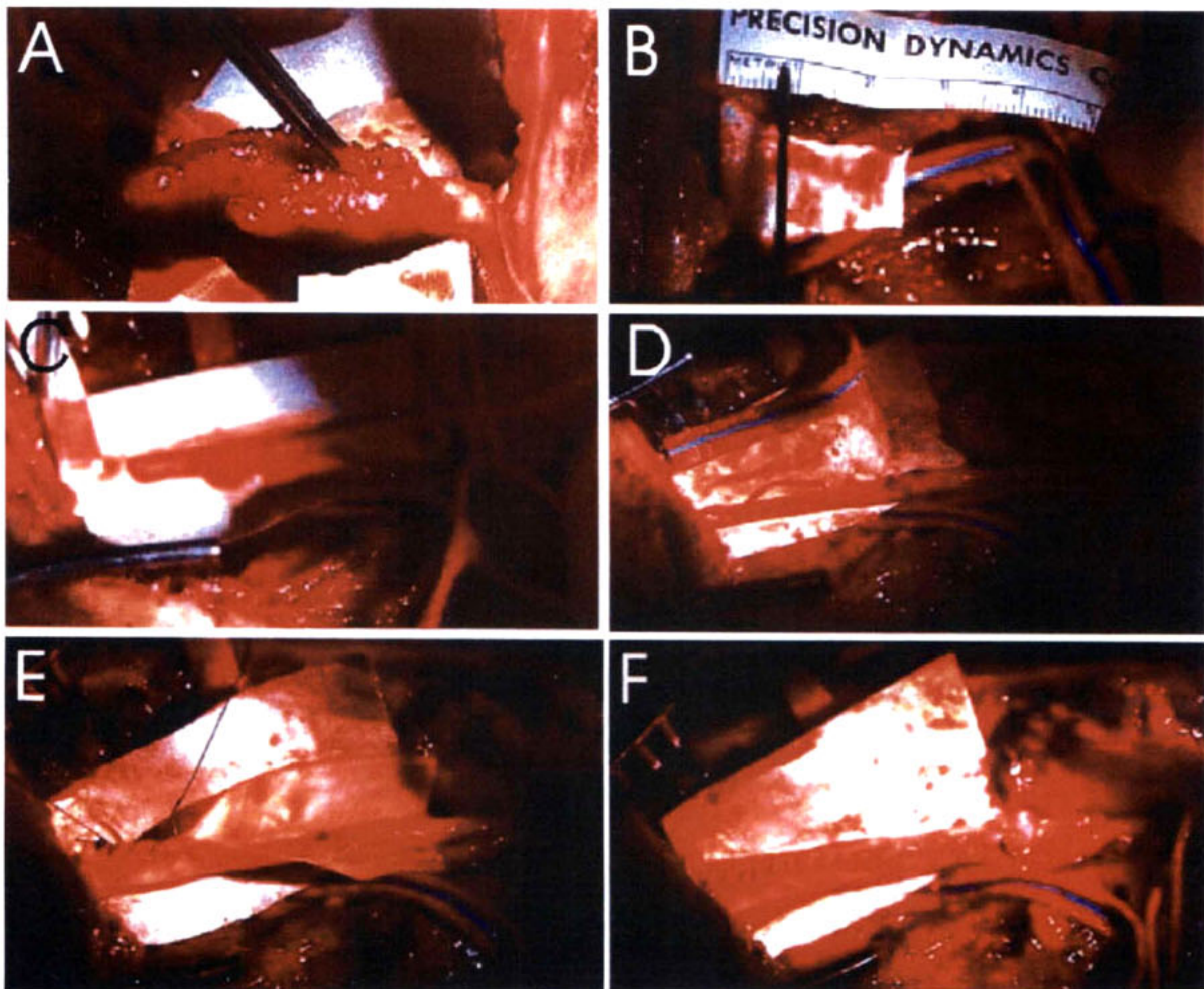


Figure 1. Photos of surgical repair procedures. A. Exposure of overlapping ends of transected radial nerve. B. Damaged nerve tissue from the proximal and distal stump are removed, resulting in a 5 cm gap. C. Positioning a length of sural nerve into the radial nerve gap. D. Two lengths of sural nerve grafted into radial nerve gap. E. Suturing a sheet of collagen around the sural nerve grafts. F. The complete nerve gap repair with the collagen tube filled with fibrin.

which concluded that he had no neurological deficits in the repaired hand. He enlisted and presently has a fulltime rigorous military job requiring full and equal use of both the repaired and un-injured hand.

Discussion

The aim of this study was to determine whether a new technique would induce more extensive neurological recovery across a 5-cm long radial nerve gap than typically induced cutaneous saphenous or sural nerve grafts. The repair involved using two sural nerve grafts surrounded by a collagen tube filled with pure fibrin.

The technique led to complete and normal motor and sensory neurological recovery that allows the patient identical use of both his repaired and un-operated hands. The observed complete sensory and motor recovery was more extensive than typically seen when sural nerve grafts are used alone to repair a nerve gap > 2 cm (16).

The study has several limitations. One is that as a case study the results are from only a single patient. Thus, further trials are required to determine the reliability of the technique in inducing neurological recovery. Another limitation is that the neurological recovery induced by sural nerve grafts is extremely variable. Therefore, it is important to consider that the observed recovery may fall within the variability of recovery induced by sural nerve grafts alone, although such complete recovery is not reported in the literature (16).

The literature indicates that good neurological recovery using only sural nerve grafts is limited to nerve gaps no longer than 2 cm in length, and that recovery decreases with increasing gap length (4, 6, 13, 16-18, 39). Further, the observed complete recovery is far greater than that seen by the surgeons involved in this study when they have repaired nerve gaps of 5-cm in length using only sural nerve grafts.

Pure fibrin within a conduit bridging a nerve gap induces neurological axon regeneration across short nerve gaps (4 mm) (40-42). However, axon regeneration is more extensive through fibrin when it is combined with trophic factors (34, 40, 43). Thus, the present data showing extensive neurological recovery most likely results from interactions of the fibrin with sural nerve-released factors.

It could be argued that tubulization of sural nerve grafts would inhibit ability to promote axon regeneration by causing the cells of the graft to become necrotic. However, the observed extensive neurological recovery indicates this is not the case.

A full clinical study is required to determine whether the nerve repair technique induces reliable neurological recovery, whether there is a maximum nerve gap length

over which the technique is effective, and whether the technique restores neurological function when applied more than 10 months post trauma, the effective limit of sural nerve grafts.

In conclusion, this study suggests that the combination of pure fibrin surrounding sural nerve grafts enhances the extent of neurological recovery induced by either pure fibrin or sural nerve grafts alone.

Acknowledgements

We wish to thank the patient for consenting to participate in this study. No conflicts of interest or economic incentives were involved in this study.

Resumen

Injertos de nervios sensoriales es el "estándar de oro" para reparar nervios periféricos con una abertura para inducir recuperación neurológica. Sin embargo, la efectividad del injerto de nervio "sural" es variable, generalmente no conduciendo a una completa recuperación sensorial y motora, y buena recuperación esta generalmente limitada a aberturas más cortas que 2 cm, y disminuyendo según aumenta el largo de los injertos. Un conducto lleno de fibrina pura conectando una abertura nerviosa conduce a solo una recuperación neurológica limitada. Examinamos la efectividad de una novedosa técnica de recuperación nerviosa en la cual una abertura nerviosa radial de 5-cm fue reparada utilizando dos injertos de nervios "surales" rodeados por un tubo de colágeno lleno de fibrina pura. El paciente tuvo una completa recuperación sensorial y motora 1 año y medio después de la operación. Concluimos que la combinación de fibrina con el injerto de nervio "sural" es responsable de inducir una extensa recuperación neurológica. Estamos examinando la técnica en una prueba clínica.

References

1. Mackinnon, SE, Kelly, L and Hunter, DA. Comparison of regeneration across a vascularized versus conventional nerve graft: case report, *Microsurgery* 1988;9:226-34.
2. Kim, DH, Kam, AC, Chandika, P, Tiel, RL and Kline, DG. Surgical management and outcome in patients with radial nerve lesions, *J Neurosurg* 2001;95:573-83.
3. Millesi, H. Peripheral nerve injuries. Nerve sutures and nerve grafting, *Scand J Plast Reconstr Surg Suppl* 1982;19:25-37.
4. Rodriguez, FJ, Gomez, N, Labrador, RO, Buti, M, Ceballos, D, Cuadras, J, et al. Improvement of regeneration with predegenerated nerve transplants in silicone chambers, *Restor Neurol Neurosci* 1999;14:65-79.
5. Meek, MF, Varejao, AS and Geuna, S. Use of skeletal muscle tissue in peripheral nerve repair: review of the literature, *Tissue Eng* 2004;10:1027-36.

6. J, IJ-P, Jansen, K, Gramsbergen, A and Meek, MF. Transection of peripheral nerves, bridging strategies and effect evaluation, *Biomaterials* 2004;25:1583-92.
7. Abernethy, DA, Rud, A and Thomas, PK. Neurotropic influence of the distal stump of transected peripheral nerve on axonal regeneration: absence of topographic specificity in adult nerve, *J Anat* 1992;180 (Pt 3):395-400.
8. Kuffler, DP. Long-distance regulation of regenerating frog axons, *J Exp Biol* 1987;132:151-60.
9. Kuffler, DP. Accurate reinnervation of motor end plates after disruption of sheath cells and muscle fibers, *J Comp Neurol* 1986;250:228-35.
10. Levi, AD, Sonntag, VK, Dickman, C, Mather, J, Li, RH, Cordoba, SC, et al. The role of cultured Schwann cell grafts in the repair of gaps within the peripheral nervous system of primates, *Exp Neurol* 1997;143:25-36.
11. Tsukahara, T, Yonekawa, Y, Tanaka, K, Ohara, O, Wantanabe, S, Kimura, T, et al. The role of brain-derived neurotrophic factor in transient forebrain ischemia in the rat brain, *Neurosurgery* 1994;34:323-31; discussion 31.
12. Nilsson, A, Dahlin, L, Lundborg, G and Kanje, M. Graft repair of a peripheral nerve without the sacrifice of a healthy donor nerve by the use of acutely dissociated autologous Schwann cells, *Scand J Plast Reconstr Surg Hand Surg* 2005;39:1-6.
13. Singh, R, Mechelse, K, Hop, WC and Braakman, R. Long-term results of transplantations to repair median, ulnar, and radial nerve lesions by a microsurgical interfascicular autogenous cable graft technique, *Surg Neurol* 1992;37:425-31.
14. Madison, RD, Archibald, SJ, Lacin, R and Krarup, C. Factors contributing to preferential motor reinnervation in the primate peripheral nervous system, *J Neurosci* 1999;19:11007-16.
15. Chiu, DT and Strauch, B. A prospective clinical evaluation of autogenous vein grafts used as a nerve conduit for distal sensory nerve defects of 3 cm or less, *Plast Reconstr Surg* 1990;86:928-34.
16. Kim, DH, Murovic, JA, Tiel, RL and Kline, DG. Management and outcomes in 318 operative common peroneal nerve lesions at the Louisiana State University Health Sciences Center, *Neurosurgery* 2004;54:1421-8; discussion 28-9.
17. Kim, DH, Connolly, SE, Gillespie, JT, Voorhies, RM and Kline, DG. Electrophysiological studies of various graft lengths and lesion lengths in repair of nerve gaps in primates, *J Neurosurg* 1991;75:440-6.
18. Navarro, X, Rodriguez, FJ, Ceballos, D and Verdu, E. Engineering an artificial nerve graft for the repair of severe nerve injuries, *Med Biol Eng Comput* 2003;41:220-6.
19. Brushart, TM, Gerber, J, Kessens, P, Chen, YG and Royall, RM. Contributions of pathway and neuron to preferential motor reinnervation, *J Neurosci* 1998;18:8674-81.
20. Mackinnon, SE and Novak, CB. Nerve transfers. New options for reconstruction following nerve injury, *Hand Clin* 1999;15:643-66, ix.
21. Hasegawa, T, Nakamura, S, Manabe, T and Mikawa, Y. Vascularized nerve grafts for the treatment of large nerve gap after severe trauma to an upper extremity, *Arch Orthop Trauma Surg* 2004;124:209-13.
22. Birch, R, Dunkerton, M, Bonney, G and Jamieson, AM. Experience with the free vascularized ulnar nerve graft in repair of supraclavicular lesions of the brachial plexus, *Clin Orthop Relat Res* 1988;96:104.
23. Mligiliche, NL, Tabata, Y and Ide, C. Nerve regeneration through biodegradable gelatin conduits in mice, *East Afr Med J* 1999;76:400-6.
24. Williams, LR. Exogenous fibrin matrix precursors stimulate the temporal progress of nerve regeneration within a silicone chamber, *Neurochem Res* 1987;12:851-60.
25. Dubovy, P and Bednarova, J. An immunocytochemical analysis of growing axons in a silicone chamber prefilled with artificial sponge matrix, *Acta Histochem* 1996;98:123-30.
26. Zhao, Q, Lundborg, G, Danielsen, N, Bjursten, LM and Dahlin, LB. Nerve regeneration in a 'pseudo-nerve' graft created in a silicone tube, *Brain Res* 1997;769:125-34.
27. Kim, D, Schallert, T, Liu, Y, Browarak, T, Nayeri, N, Tessler, A, et al. Transplantation of genetically modified fibroblasts expressing BDNF in adult rats with a subtotal hemisection improves specific motor and sensory functions, *Neurorehabil Neural Repair* 2001;15:141-50.
28. Anderson, PN and Turmaine, M. Peripheral nerve regeneration through grafts of living and freeze-dried CNS tissue, *Neuropathol Appl Neurobiol* 1986;12:389-99.
29. Archibald, SJ, Krarup, C, Shefner, J, Li, ST and Madison, RD. A collagen-based nerve guide conduit for peripheral nerve repair: an electrophysiological study of nerve regeneration in rodents and nonhuman primates, *J Comp Neurol* 1991;306:685-96.
30. Mears, S, Schachner, M and Brushart, TM. Antibodies to myelin-associated glycoprotein accelerate preferential motor reinnervation, *J Peripher Nerv Syst* 2003;8:91-9.
31. Dahlin, LB. Stimulation of regeneration of the sciatic nerve by experimentally induced inflammation in rats, *Scand J Plast Reconstr Surg Hand Surg* 1992;26:121-5.
32. Bertleff, MJ, Meek, MF and Nicolai, JP. A prospective clinical evaluation of biodegradable neurolac nerve guides for sensory nerve repair in the hand, *J Hand Surg [Am]* 2005;30:513-8.
33. Baier, H and Bonhoeffer, F. Axon guidance by gradients of a target-derived component, *Science* 1992;255:472-5.
34. Galla, TJ, Vedecnik, SV, Halbgewachs, J, Steinmann, S, Friedrich, C and Stark, GB. Fibrin/Schwann cell matrix in poly-epsilon-caprolactone conduits enhances guided nerve regeneration, *Int J Artif Organs* 2004;27:127-36.
35. Lee, DY, Choi, BH, Park, JH, Zhu, SJ, Kim, BY, Huh, JY, et al. Nerve regeneration with the use of a poly(l-lactide-co-glycolic acid)-coated collagen tube filled with collagen gel, *J Craniomaxillofac Surg* 2006;34:50-6.
36. Wu, S, Suzuki, Y, Tanihara, M, Ohnishi, K, Endo, K and Nishimura, Y. Repair of facial nerve with alginate sponge without suturing: an experimental study in cats, *Scand J Plast Reconstr Surg Hand Surg* 2002;36:135-40.
37. Fansa, H, Keilhoff, G, Wolf, G and Schneider, W. Tissue engineering of peripheral nerves: A comparison of venous and acellular muscle grafts with cultured Schwann cells, *Plast Reconstr Surg* 2001;107:485-94; discussion 95-6.
38. Azhar, MM and Sara, TA. Comparison of nerve graft and artificial conduits for bridging nerve defects, *Med J Malaysia* 2004;59:578-84.
39. Samardzic, MM, Rasulic, LG and Grujicic, DM. Results of cable graft technique in repair of large nerve trunk lesions, *Acta Neurochir (Wien)* 1998;140:1177-82.
40. Pittier, R, Sauthier, F, Hubbell, JA and Hall, H. Neurite extension and in vitro myelination within three-dimensional modified fibrin matrices, *J Neurobiol* 2005;63:1-14.
41. Palazzi, S, Vila-Torres, J and Lorenzo, JC. Fibrin glue is a sealant and not a nerve barrier, *J Reconstr Microsurg* 1995;11:135-9.
42. Zeng, L, Huck, S, Redl, H and Schlag, G. Fibrin sealant matrix supports outgrowth of peripheral sensory axons, *Scand J Plast Reconstr Surg Hand Surg* 1995;29:199-204.
43. Tsai, EC, Dalton, PD, Shoichet, MS and Tator, CH. Matrix inclusion within synthetic hydrogel guidance channels improves specific supraspinal and local axonal regeneration after complete spinal cord transection, *Biomaterials* 2006;27:519-33.